

Complete genome sequences of four novel human gammapapillomavirus types, HPV-219, HPV-220, HPV-221, and HPV-222, isolated from penile skin swabs from South African men

Item Type	Article
Authors	Murahwa, A.T;Meiring, T.L;Mbulawa, Z.Z.A;Williamson, A.L
Citation	Murahwa AT, Meiring TL, Mbulawa ZZA, Williamson AL. Complete Genome Sequences of Four Novel Human Gammapapillomavirus Types, HPV-219, HPV-220, HPV-221, and HPV-222, Isolated from Penile Skin Swabs from South African Men. Genome Announc. 2018 Jun 21;6(25):e00584-18. doi: 10.1128/genomeA.00584-18.
DOI	https://doi.org/10.1128/genomea.00584-18
Publisher	American Society for Microbiology
Journal	Genome Announcements
Rights	Attribution 3.0 United States
Download date	2024-08-07 17:53:39
Item License	http://creativecommons.org/licenses/by/3.0/us/
Link to Item	https://pubmed.ncbi.nlm.nih.gov/29930074/



Complete Genome Sequences of Four Novel Human *Gammapapillomavirus* Types, HPV-219, HPV-220, HPV-221, and HPV-222, Isolated from Penile Skin Swabs from South African Men

 Alltalents T. Murahwa,^a Tracy L. Meiring,^a Zizipho Z. A. Mbulawa,^{a,b} Anna-Lise Williamson^{a,c}

^aDivision of Medical Virology, Department of Pathology and Institute of Infectious Diseases & Molecular Medicine, Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa

^bCenter for HIV and STIs, National Institute for Communicable Disease, National Health Laboratory Service, Johannesburg, South Africa

^cSAMRC Gynaecological Cancer Research Centre, Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa

ABSTRACT Four novel human gammapapillomaviruses were characterized from penile specimens using genome amplification, cloning, and sequencing. The HPV-219 L1 gene showed 87% nucleotide identity to that of HPV-213 of species gamma-13, HPV-220 had 72% identity to L1 of HPV-212 (gamma-17), HPV-221 had 80% identity to L1 of HPV-142 (gamma-10), and HPV-222 had 73% nucleotide identity to L1 of HPV-162 (gamma-19).

Human papillomaviruses (HPVs) are small nonenveloped DNA viruses of the *Papillomaviridae* family that infect mucosal and cutaneous epithelia (1, 2). The genus *Gammapapillomavirus* is the most divergent and rapidly growing genus of the family, with 27 species and 98 officially recognized genotypes (3). Here, we describe the characterization of four novel gammapapillomaviruses initially discovered using deep sequencing of the HPV L1 FAP amplicon region (4).

The penile swab collection and DNA extraction procedures have been described previously (5). The full genomes were amplified as single amplicons with back-to-back primers based on the FAP amplicon sequence and the LongRange HotStart PCR kit (Kapa Biosystems, USA). PCR products cloned into the TOPO XL vector (Thermo Fisher, USA) were sequenced on the Illumina MiSeq (2 × 300 bp) by Macrogen, Inc. (South Korea). Genome assembly was done using the *de novo* assembly function in CLC Genomics Workbench (GW) version 8.5.1 (Qiagen, USA). Splice site prediction was carried out as outlined by Van Doorslaer et al. (6). Reference clones of HPV-219 (7,108 bp), HPV-220 (7,381 bp), HPV-221 (7,326 bp), and HPV-222 (7,275 bp) were sent to the International HPV Reference Center (http://www.nordicehealth.se/hpvcenter/reference_clones/) for confirmation and assignment of type numbers.

HPV-219 is phylogenetically most closely related to HPV-213 of the gamma-13 species, sharing 87% identity in the L1 gene, HPV-220 is most closely related to HPV-212 (72% L1 identity) of gamma-17, HPV-221 is most closely related to HPV-142 (80% L1 identity) of gamma-10, and HPV-222 is most closely related to HPV-162 (73% L1 identity) of gamma-19. These HPV genotypes share <90% identity in the L1 gene (1); therefore, all four viruses are novel genotypes. The genomic organization was typical of gammapapillomaviruses, encoding five early (E1, E2, E4, E6, and E7) and two late (L1 and L2) proteins, and lacking the E5 gene. HPV-221 and HPV-222 did not have start codons for the E4 gene, but the spliced E1[^]E4 transcript, which encodes the primary E4 gene product (7), was identified in all four genomes.

Received 24 May 2018 Accepted 24 May 2018 Published 21 June 2018

Citation Murahwa AT, Meiring TL, Mbulawa ZZA, Williamson A-L. 2018. Complete genome sequences of four novel human *Gammapapillomavirus* types, HPV-219, HPV-220, HPV-221, and HPV-222, isolated from penile skin swabs from South African men. *Genome Announc* 6:e00584-18. <https://doi.org/10.1128/genomeA.00584-18>.

Copyright © 2018 Murahwa et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Anna-Lise Williamson, anna-lise.williamson@uct.ac.za.

All four viruses had a TATA box (TATAAA) (8) and palindromic E2-binding sites (ACC-N₆-GGT) in their long control regions (9). An ATP binding site, G(x)₄GK(T/S) (10, 11), was present in the C-terminal region of the E1 proteins of the viruses. Two conserved zinc binding domains [CxxC(x)₂₉CxxC] (12) were identified in the E6 proteins and one in the E7 protein of all the viruses. HPV-219 additionally contained a putative PDZ binding domain, x(T/S)x(L/V) (13), in the E6 N-terminal region.

To conclude, we discovered four novel HPV genotypes of the *Gammapapillomavirus* genus. This knowledge expands the heterogeneity of the ever-growing members of the gammapapillomaviruses. The prevalence and clinical importance of these novel gammapapillomaviruses warrant further investigation.

Accession number(s). The GenBank accession numbers for the HPV-219, HPV-220, HPV-221, and HPV-222 genome sequences are [MH172376](#), [MH172377](#), [MH172378](#), and [MH172379](#), respectively.

ACKNOWLEDGMENTS

We thank J. Dillner, C. Eklund, and D. Bzhalava from the International HPV Reference Center for receiving and confirming the novel HPV types.

A.T.M. is the recipient of a Ph.D. fellowship from the Letten Research Foundation of Norway, Oslo, administered by B. Stray-Pedersen. This study was supported in part by the Poliomyelitis Research Fund (PRF) of South Africa and the National Research Fund (NRF) of South Africa.

This work is based upon research supported by the South African Research Chairs Initiative of the Department of Science and Technology and the NRF.

REFERENCES

- Bernard H-U, Burk RD, Chen Z, van Doorslaer K, Zur Hausen H, de Villiers E-M. 2010. Classification of papillomaviruses (PVs) based on 189 PV types and proposal of taxonomic amendments. *Virology* 401:70–79. <https://doi.org/10.1016/j.virol.2010.02.002>.
- de Villiers E-M, Fauquet C, Broker TR, Bernard H-U, Zur Hausen H. 2004. Classification of papillomaviruses. *Virology* 324:17–27. <https://doi.org/10.1016/j.virol.2004.03.033>.
- Mühr LSA, Eklund C, Dillner J. 2018. Towards quality and order in human papillomavirus research. *Virology* 519:74–76. <https://doi.org/10.1016/j.virol.2018.04.003>.
- Meiring TL, Mbulawa ZZA, Lesosky M, Coetzee D, Williamson A-L. 2017. High diversity of alpha, beta and gamma human papillomaviruses in genital samples from HIV-negative and HIV-positive heterosexual South African men. *Papillomavirus Res* 3:160–167. <https://doi.org/10.1016/j.pvr.2017.05.001>.
- Mbulawa ZZ, Coetzee D, Marais DJ, Kamupira M, Zwane E, Allan B, Constant D, Moodley JR, Hoffman M, Williamson A-L. 2009. Genital human papillomavirus prevalence and human papillomavirus concordance in heterosexual couples are positively associated with human immunodeficiency virus coinfection. *J Infect Dis* 199:1514–1524. <https://doi.org/10.1086/598220>.
- Van Doorslaer K, Li Z, Xirasagar S, Maes P, Kaminsky D, Liou D, Sun Q, Kaur R, Huyen Y, McBride AA. 2017. The Papillomavirus Episteme: a major update to the papillomavirus sequence database. *Nucleic Acids Res* 45:D499–D506. <https://doi.org/10.1093/nar/gkw879>.
- Doorbar J. 2013. The E4 protein; structure, function and patterns of expression. *Virology* 445:80–98. <https://doi.org/10.1016/j.virol.2013.07.008>.
- de Villiers E-M, Gunst K. 2009. Characterization of seven novel human papillomavirus types isolated from cutaneous tissue, but also present in mucosal lesions. *J Gen Virol* 90:1999–2004. <https://doi.org/10.1099/vir.0.011478-0>.
- Newhouse CD, Silverstein SJ. 2001. Orientation of a novel DNA binding site affects human papillomavirus-mediated transcription and replication. *J Virol* 75:1722–1735. <https://doi.org/10.1128/JVI.75.4.1722-1735.2001>.
- McBride AA. 2008. Replication and partitioning of papillomavirus genomes. *Adv Virus Res* 72:155–205. [https://doi.org/10.1016/S0065-3527\(08\)00404-1](https://doi.org/10.1016/S0065-3527(08)00404-1).
- Titolo S, Pelletier A, Sauvé F, Brault K, Wardrop E, White PW, Amin A, Cordingley MG, Archambault J. 1999. Role of the ATP-binding domain of the human papillomavirus type 11 E1 helicase in E2-dependent binding to the origin. *J Virol* 73:5282–5293.
- Wayengera M. 2012. Zinc finger arrays binding human papillomavirus types 16 and 18 genomic DNA: precursors of gene-therapeutics for *in-situ* reversal of associated cervical neoplasia. *Theor Biol Med Model* 9:30. <https://doi.org/10.1186/1742-4682-9-30>.
- Bolatti EM, Chouhy D, Casal PE, Pérez GR, Stella EJ, Sanchez A, Gorosito M, Bussy RF, Giri AA. 2016. Characterization of novel human papillomavirus types 157, 158 and 205 from healthy skin and recombination analysis in genus γ -Papillomavirus. *Infect Genet Evol* 42:20–29. <https://doi.org/10.1016/j.meegid.2016.04.018>.